



Nathan Jahnke, Brian E. Whipker, John Dole, Mike Munster, and Inga Meadows¹

Lantana: *Diagnosing Lower Leaf Angular Spots*

When one observes lower leaf angular spots on lantana, there are three potential causal pathogens: foliar nematodes, bacteria, or downy mildew. Foliar nematodes are easy to self-diagnose by following the steps outlined in this Alert. [Foliar Nematode Video Link](#)

When leaves start to develop angular leaf spots, there are three potential casual pathogens: foliar nematodes, bacteria, or downy mildew (see sidebar for discussion of mimics on page 5).

In this situation, the pictured lantana (Fig. 1) received a confirmed diagnosis of foliar nematodes from an experienced pathologist. It contained so many nematodes it surprised him! Usually we think of nematodes as a pathogen that attacks roots, but the *Aphelenchoides* spp. are unique. They feed on leaf tissue in the mesophyll. They do require a film water to move which means that they can be easily spread by splashing water from leaf to leaf. Typically, stomata are the main points of entry into leaves where they feed on leaf tissue in the mesophyll. New nematodes develop from larva which hatch from eggs laid inside the leaf tissue. Dried plant debris also can harbor dormant nematodes allowing them wait for moisture and new plant material to infect. As they feed on leaf tissue, symptoms begin as a patchy yellowing

¹ Nathan Jahnke is a M.S. student in the Dept. of Horticultural Science at NC State University, Brian E. Whipker and John Dole are Professors of Floriculture, Mike Munster is with the NCSU Plant Disease and Insect Clinic, and Inga Meadows is the Extension Vegetable and Floriculture Pathologist.

Contact Info: bwhipker@ncsu.edu

2016 Sponsors



Figure 1. Angular spots appear on the lower foliage of lantana due to a foliar nematode infestation.

Photo by Brian Whipker

e-GRO Alert

www.e-gro.org

CONTRIBUTORS

Dr. Nora Catlin

Floriculture Specialist
Cornell Cooperative Extension - Suffolk County
nora.catlin@cornell.edu

Dr. Chris Currey

Assistant Professor of Floriculture
Iowa State University
ccurrey@iastate.edu

Thomas Ford

Commercial Horticulture Educator
Penn State Extension
tgf2@psu.edu

Dan Gilrein

Entomology Specialist
Cornell Cooperative Extension - Suffolk County
dog1@cornell.edu

Dr. Joyce Latimer

Floriculture Extension & Research
Virginia Tech
jltime@vt.edu

Dr. Roberto Lopez

Floriculture Extension & Research
Purdue University
rglopez@purdue.edu

Dr. Neil Mattson

Greenhouse Research & Extension
Cornell University
neil.mattson@cornell.edu

Dr. Rosa E. Raudales

Greenhouse Extension Specialist
University of Connecticut
rosa.raudales@uconn.edu

Dr. Beth Scheckelhoff

Ext. Educator – Greenhouse Systems
The Ohio State University
scheckelhoff.11@osu.edu

Lee Stivers

Extension Educator – Horticulture
Penn State Extension, Washington County
ljs32@psu.edu

Dr. Paul Thomas

Floriculture Extension & Research
University of Georgia
pathomas@uga.edu

Dr. Brian Whipker

Floriculture Extension & Research
NC State University
bwhipker@ncsu.edu

Heidi Wollaeger

Floriculture Outreach Specialist
Michigan State University
wollaeger@anr.msu.edu

Copyright © 2016

Where trade names, proprietary products, or specific equipment are listed, no discrimination is intended and no endorsement, guarantee or warranty is implied by the authors, universities or associations.

(Fig. 2) which can develop angular necrosis (Fig. 3) between veins. Plants can be asymptomatic, showing no symptoms, and still contain a population of nematodes.

The host range for foliar nematodes is very large. Bedding plants with reported infections include: ageratum, antirrhinum (snapdragon), argyranthemum, begonia, chrysanthemum, dahlia, ferns, gerbera, gomphrena, helichrysum, hellebore, hosta, lantana, mimulus, pentas, petunia, salvia, solenostemon (coleus), strobilanthes, tithonia, torenia, verbena, and zinnia. (For a complete listing of plants with confirmed infestations by *Aphelenchoides* spp., see L.M. Kohl reference.)

Diagnosis

According to Shurtleff and Averre (2000), foliar nematodes are colorless, microscopic roundworms that are 0.5 to 1.2 mm long. They reside in the mesophyll region of leaves. Leaf veins are natural barriers to their movement which leads to the development of angular necrotic spots between veins.

You can send samples into a diagnostic clinic or check for nematodes yourself with a dissecting or compound microscope, with the capability of magnifying up to 40X.



Figure 2. Initial sign of a foliar nematode infestation is the patchy, angular spots on the lower leaves.

Photo by Brian Whipker



Figure 3. Necrotic spotting develops as the leaf tissue dies.
Photo by Brian Whipker

Proceed with the following steps:

- 1) Remove a diseased leaf from a plant that contains healthy and necrotic portions.
- 2) Cut out a small square with a knife or scalpel. Make sure that the sample contains part of the healthy and diseased tissue, as this is where the nematodes would be located.
- 3) Cut that small square into 1 cm wide strips.
- 4) Place the cut tissue on a glass slide with 1 - 2 drops of water.
- 5) Place a cover slip over the tissue.
- 6) Allow the cut edges to sit in water for at least 5 minutes to allow time for nematodes to migrate out of the leaf tissue.

Cooperating Universities

UConn



Cornell University



The University of Georgia

IOWA STATE UNIVERSITY

MICHIGAN STATE
UNIVERSITY

NC STATE



THE OHIO STATE
UNIVERSITY

PENNSTATE



Cooperative Extension
College of Agricultural Sciences

PURDUE
UNIVERSITY



VirginiaTech
Invent the Future

MAUMEE VALLEY GROWERS
Choose the Very Best.



In cooperation with our local and
state greenhouse organizations



Indiana
FLOWER
GROWERS
Association



Michigan Floriculture Growers Council



CONNECTICUT
GREENHOUSE
GROWERS
ASSOCIATION

7) Place the sample under a dissecting or compound microscope and starting with the lowest magnification, search along the leaf tissue for the 0.5 - 1.2 mm roundworm (Fig. 4).

8) Increase the magnification to search in more detail, starting at the cut edges. *Aphelenchoides* and any other plant-parasitic nematode will have a small needle-like structure called a stylet in the anterior portion.

Management

Take the following steps to manage foliar nematodes:

- Discard infected plant material and soil or soilless media.
- Clean and then disinfest or steam sterilize pots before re-using.

- Quarantine plants that were growing nearest to infected plants.
- Remove plant debris from propagation and production areas.
- Remove weeds from greenhouse floors, benches and pots.
- Avoid overhead irrigation and splashing water on leaf surfaces.
- Increase plant spacing, if possible, to favor air movement and rapid drying.
- Do not handle plants when foliage is wet.

The described cultural practices are the best methods of control along with using clean stock, since there are no effective chemicals that can eliminate foliar nematodes. Scout held-over stock plants and

the common “pet” plants that are brought into the greenhouse. When receiving new plant material, inspect, and remove infected product promptly.

Literature Cited

Kohl, L.M., 2008. Population dynamics and dispersal gradient of *Aphelenchoides fragariae* in the woody ornamental Lantana camera. N. Carolina State Univ., Raleigh, MS Thesis. (<http://www.lib.ncsu.edu/resolver/1840.16/1457>) For host listing, please refer to pages 22-58.

Shurtleff, M.C. and Averre, C.W. 2000. Diagnosing Plant Diseases Caused by Nematodes. APS Press. St. Paul, Minnesota, USA. 187pp.



Figure 4. A foliar nematode emerging from the cut surface of a leaf, under >40X magnification. Photo by Mike Munster

Mimics

The second problem which results in angular lesions is bacterial leaf spot. (If one counts the number of samples that are submitted to the NCSU Plant Disease and Insect Clinic, bacterial leaf spot problems are more common than foliar nematode samples.) Bacteria such as *Acidovorax*, *Pseudomonas*, and *Xanthomonas* can cause these water soaked, angular lesions which can also develop yellow halos (Fig. 5).



Figure 5. A bacterial infection of alopecia.
Photo by Brian Whipker

Finally, an infection by the downy mildew ifungus also can result in angular yellowing and necrosis. This symptomology is especially true for Buddleia, *Buddleja davidii*) (Fig. 6). Other ornamentals with similar symptomology include rose, veronica, and agastache (Fig. 7).



Figure 6. A downy mildew infection of buddleia.
Photo by Mike Munster



Figure 7. A downy mildew infection of agastache.
Photo by Mike Munster

Greenhouse plants with reported infestations by foliar nematodes (*Aphelenchoides fragariae*) [partial list].

| | | |
|-----------------------|--------------------|--------------------------|
| African Violet | Cyclamen | Lilium |
| Anemone | Dahlia | Mimulus |
| Anthurium | Delphinium | Pentas |
| Argyranthemum | Foxglove | Petunia |
| Aster | Geranium | Phlox, Creeping |
| Azalea | Gerbera | Primula |
| Baptisia | Gloxinia | Ranunculus |
| Basil | Gomphrena | Rudbeckia |
| Begonia | Helichrysum | Salvia |
| Boston Fern | Hellebore | Snapdragon (antirrhinum) |
| Bird's Nest Fern | Heuchera | Strobilanthes |
| Cactus, Thanksgiving | Hibiscus | Strawberry |
| Calceolaria | Hosta | Tithonia |
| Chrysanthemum | Hydrangea | Torenia |
| Cineraria | Impatiens | Verbena |
| Coleus (solenostemon) | Indian Rubber Tree | Zinnia |
| Columbine (Aquilegia) | Iris | |
| Crossandra | Lantana | |